Human Diffuse Large B-Cell Lymphoma Cells (OCI-LY3)

Catalog Number: RWT-492

Product Description

The OCI-LY3 cell line was established in 1983 from the bone marrow of a 52-year-old male diagnosed with B-cell non-Hodgkin lymphoma (diffuse large B-cell lymphoma, DLBCL, stage IVB) at relapse. Literature reports indicate that these cells are EBV-negative and lack the classical t(8;14) translocation.

Product Characteristics

- **Tissue Source:** Human bone marrow
- **Growth Pattern:** Round cells growing singly or in clusters; mixed suspension and adherent growth
- **Cell Quantity:** >1 × 10^6 cells per vial (1 mL)
- **Contamination Testing:** Negative for mycoplasma, bacteria, yeast, and fungi
- **Packaging:** 1 mL frozen cell suspension in cryovial

Shipping & Storage

- Shipped on **dry ice** (cryopreserved)

Upon Receipt: - Immediately transfer the vial to **liquid nitrogen storage**, or - Begin cell culture immediately

Important: - Take photos upon receipt - If contamination is observed within 3 days, document with photos and contact us immediately

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Product Use

⚠️ **For Research Use Only**
Not intended for human or animal use. Not approved for diagnostic applications.

Cell Culture Instructions

1. Preparation

Prepare complete culture medium: - IMDM: 79% - Fetal Bovine Serum (FBS): 20% - Penicillin-Streptomycin (P/S): 1%

Culture Conditions: - Temperature: 37°C - CO_2: 5% - Air: 95% - Humidity: 70–80%

Cryopreservation Medium: - 90% FBS + 10% DMSO (prepare fresh before use)

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2. Cell Thawing & Initiation

1. Rapidly thaw the cryovial in a 37°C water bath
2. Add 4 mL culture medium and gently resuspend
3. Centrifuge at 1000 rpm for 4 minutes
4. Discard supernatant and resuspend in 1–2 mL medium
5. Transfer to culture flask and incubate overnight
6. Replace medium the next day and monitor cell density

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3. Subculture Procedure

Subculture when cells reach **80–90% confluence**:

1. Remove medium and wash cells 1–2 times with PBS (without Ca^{2+}/Mg^{2+})
2. Add 2 mL 0.25% Trypsin-0.53 mM EDTA and incubate at 37°C for 1–2 minutes
3. Observe under microscope; once cells round up and detach:
4. Stop digestion with culture medium
5. Add 6–8 mL medium, resuspend gently
6. Centrifuge at 1000 rpm for 4 minutes
7. Resuspend in 1–2 mL medium

Passaging Ratio: - First passage: 1:2 (recommended; freeze backup) - Routine passage: 1:2 to 1:5 depending on growth

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4. Cryopreservation

Proceed when cells are in optimal growth condition.

Example (T25 Flask):
1. Remove medium and wash with PBS 2. Add 1 mL trypsin and detach cells 3.Neutralize with 2 mL complete medium 4. Count cells using a hemocytometer 5. Centrifuge at 1000 rpm for 5 minutes 6. Resuspend in FBS and add DMSO to final concentration of 10% 7. Aliquot 1 mL per cryovial (>1×10^6 cells/vial) 8. Place at -80°C, then transfer to liquid nitrogen after ≥2 hours 9. Record storage location

Important Notes

1. Contact us immediately if:
2. Dry ice has completely evaporated
3. Cryovial is damaged or open
4. Contamination is observed

Biosafety Notice:

5. All human/animal cells are potentially hazardous
6. Follow Biosafety Level 1 (BSL-1) guidelines
7. Wear appropriate PPE (gloves, lab coat, eye protection)
8. Sterilize all waste before disposal

Company Information

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